# Mechanisms of Experimental Cancer Cachexia

## Local Involvement of IL-1 in Colon-26 Tumor

# Gideon Strassmann,1\* Yoshihiro Masui,† Richard Chizzonite,‡ and Miranda Fong\*

\*Department of Immunology, Otsuka America Pharmaceutical Inc., Rockville, MD 20850; †the Cellular Technology Institute, Otsuka Pharmaceutical Co., Ltd., Tokushima, Japan 771-01; and the †Department of Molecular Genetics, Hoffman-LaRoche Inc., Nutley, NJ 07110

ABSTRACT. In the colon-26 (C-26) tumor model, the cytokine IL-6 is an important factor involved in experimental cancer cachexia. Recent in vitro data indicated that IL-1 plays a role in the interaction between host macrophages and C-26 cells that express IL-1R, resulting in the amplification of tumor IL-6 production. To investigate the role of IL-1 on the development of C-26 cachexia in vivo, the effect of specific blockade of the action of IL-1 with reagents against IL-1R was evaluated. Both IL-1R antagonist (IL-1RA) and the mAb 35F5 directed against IL-1R type I, prevented binding of radioactive IL-1, and inhibited IL-1-induced IL-6 synthesis by the C-26 cell line. Whereas a systemic administration of these reagents did not reverse weight loss in C-26-bearing mice, intratumoral injections of IL-1RA significantly reduced cachexia. Furthermore, body composition analysis confirmed that this treatment improved lean tissue and fat, as well as hypoglycemia and serum IL-6 level. The fact that the treatment did not change the tumor burden suggests that it affected the host directly. These results support the hypothesis that, at the microenvironment of the C-26 tumor, IL-1 is involved in the cachexia endured by the host. *Journal of Immunology*, 1993, 150: 2341.

eoplastic diseases are frequently associated with metabolic changes collectively known as cancer cachexia. These abnormalities include wasting of both fat and muscle tissues, anorexia, asthenia, hypoglycemia, and anemia (1, 2). Cachexia long has been recognized as an important cause of death in cancer patients (3), and patients who exhibit weight loss have a reduced response to chemotherapy (4). Understanding the mechanisms that lead to cachexia therefore is important.

TNF has been suggested as a mediator of cancer cachexia because it suppresses key metabolic enzymes and induces anorexia and weight loss in animals (5–8). Recently, however, an experimental cachexia model has been identified that appears to involve another cytokine. The model uses

a cell line derived from C-26,<sup>2</sup> which retains the transplantability of the original tumor in syngeneic mice, and fulfills the criteria of early onset wasting without apparent anorexia. It also involves a relatively small tumor burden (9). In at least this model, IL-6 appears to have a more significant role than TNF in mediating the myriad parameters of cachexia (9). In culture, the C-26.IVX cell line expresses high affinity IL-1R type I, and fM concentrations of IL-1 but not TNF, upregulates IL-6 production (10). In addition, significant potentiation of tumor IL-6 secretion can be seen when the line is co-cultured with syngeneic mononuclear phagocytes. In this cellular interaction, the mAb 35F5, directed against murine IL-1R type I, blocks IL-6 synthesis (10).

IL-1RA is a naturally occurring protein (11), which is useful in blocking several IL-1-mediated pathologies in vivo (12). Accordingly, utilizing IL-1R blocking reagents, the role of IL-1 in the development of C-26-mediated cachexia was investigated.

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<sup>&</sup>lt;sup>1</sup> Address correspondence and reprint requests to Dr. Gideon Strassmann, Department of Immunology, Otsuka America Pharmaceutical Inc., 9900 Medical Center Drive, Rockville, Maryland, 20850.

 $<sup>^2</sup>$  Abbreviations used in this paper: C-26, colon-26 adenocarcinoma; IL-1RA, IL-1R antagonist; (CD)F<sub>1</sub>, BALB/c  $\times$  DBA/2F<sub>1</sub>.

#### Materials and Methods

Mice

Virus-free, male BALB/C  $\times$  DBA/2 (CD)F<sub>1</sub>, mice were purchased from Charles River Breeding Laboratories (Wilmington, MA). Mice were housed under conventional conditions and used at 10 to 12 wk of age.

#### Reagents

Human rIL-1β was a gift from Dr. Y. Hirai (Otsuka Pharmaceutical, Tokushima, Japan). [125I]IL-1α (sp.act. 2000 Ci/mmol.) was from Amersham (Arlington Heights, IL). Purified IL-1RA was obtained after the cloning, expression, and purification procedures described previously (11, 13). The sterile material had a purity of greater than 99% by SDS-PAGE, and endotoxin content was determined to be less than 0.5 ng/mg of protein. For in vivo experiments, IL-1RA was diluted in sterile Dulbecco's Ca<sup>++</sup>- and Mg<sup>++</sup>-free PBS (GIBCO, Grand Island, NY). This diluent also served as a vehicle for control injections. The rat mAb 35F5 (14) (IgG1) was obtained from Dr. R. Chizzonite (Hoffman-LaRoche, Nutley, NJ). Purified rat IgG was obtained from Sigma (St. Louis, MO).

#### **Assays**

The IL-1 radioreceptor assay on C-26.IVX (obtained without trypsinization) and on EL-4.6.1 cells was performed as previously described (10). The presence of IL-6 in serum and culture-conditioned medium was performed by utilizing the B-9 cell line assay (10) and ELISA (Endogen, Boston, MA). The addition of mAb against murine IL-6 and a murine IL-6R completely abrogated IL-6-dependent proliferation by test samples.

#### Measurement of cachexia markers

Mice were inoculated with  $0.5 \times 10^6$  C-26.IVX cells s.c. to the right flank as described (9, 10). Treatments were performed as indicated in the tables. Mice were weighed between 9 a.m. and 11 a.m. three times per week. The length and width of their tumors were measured by using an engineering caliper, and estimation of tumor weight was calculated, as described, for the same tumor (15). Significant weight loss in C-26-bearing mice occurred between 12 and 14 days after tumor inoculation. Host weight was calculated by subtracting tumor weight (obtained by resection) from total weight. Blood was obtained by cardiac puncture (approximately 0.8 ml), and serum was harvested after the clotting of blood at room temperature for 1 h. Serum was kept frozen (-45°C) until analysis. Measurements of serum glucose were performed using an Ektachem DT-60 analyzer (Eastman Kodak Co., Rochester, NY). Dry weight was determined (after removal of the tumor, blood, and right epididymal fat) by oven drying for 3 days at 85°C.

Table | IL-1RA and anti-IL-1R type | mAb block binding of radioactive | IL-1 to C-26.IVX cells

Addition to Padiorecentor Assault	[125]]IL-1α Bound (cpm)b		
Addition to Radioreceptor Assay <sup>a</sup>	C-26.IVX	EL-4.6.1	
Medium	2237 ± 10	3483 ± 25	
Human IL-1β (100 ng/ml)	$400 \pm 10$	$491 \pm 5$	
Rat IgG (2 µg/ml)	$2257 \pm 80$	$3338 \pm 210$	
Anti-IL-1R (2 µg/ml)	$495 \pm 5$	$457 \pm 35$	
IL-1RA (30 ng/ml)	$437 \pm 20$	$400 \pm 30$	

<sup>&</sup>lt;sup>a</sup> Radioreceptor assay was performed as described in *Materials and Methods*.

#### Statistical analysis

Results are presented as mean  $\pm$  SD. Differences in cachexia markers were calculated using computerized analysis of variance.

#### **Results and Discussion**

First we determined whether reagents capable of recognizing IL-1R would block binding of radioactive IL-1 to the C-26.IVX cell line. As shown in Table I, the mAb 35F5 and IL-1RA completely blocked binding of [ $^{125}$ I]IL-1 $\alpha$  to both the C-26 cell line and to EL-4.6.1 cells in a radioreceptor assay. In a dose-dependent manner, both the mAb and the IL-1RA inhibited IL-1 induced IL-6 production by the tumor line (Table II). On a molar basis, IL-1RA was approximately 240-fold more active than the 35F5 mAb in inhibiting IL-6 synthesis. Also in the table, IL-1RA at millimolar concentration had no agonist activity in inducing IL-6 production by the tumor line.

Next we examined the role of IL-1 in C-26 mediated cachexia. Mice were inoculated with the tumor line and treated with the mAb and IL-1RA, as indicated in Table III. No significant or sustained improvement in tumormediated weight loss could be seen when these reagents were administered systemically. Of note, the amount of IL-1RA used in this experiment exceeded the amount used to protect mice from graft-vs-host disease mortality (16) and to block IL-1 induction of IL-6 in vivo (17). The amount of 35F5 mAb used was greater than that shown to increase mortality of irradiated mice (18) and to attenuate turpentine-induced weight loss (19). Together with our previously published data (9), these results suggest that IL-1 does not act in concert with IL-6 in the circulation of C-26 bearing mice as they were losing weight. Nevertheless, the possibility still exists that the amount of blocking reagents used was insufficient to reach the tumor and effectively exert their action. Thus, we attempted to prevent cachexia by the direct administration of IL-1RA to the tumor. As shown in Tables IV and V, intratumoral injections of IL-1RA significantly improved the deleterious systemic effects of the tumor. Typical cachectic parameters, including

<sup>&</sup>lt;sup>b</sup> Results are expressed as cpm bound  $\pm$  SD.

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Table II IL-1RA and anti-IL-1R type I mAb inhibit IL-1-induced IL-6 production by C-26.IVX cells<sup>a</sup>

Addition to Culture		IL-6 Production			
IL-1 IL-1RA	A-4: N 1D	Bioassay		ELISA	
	IL-TKA	Anti-JL-1R	24 h	48 h	48 h
_	-		3	35	30
+	_	-	330	1100	1120
+	100	_	6	50	50
+	30	_	10	87	$NT^b$
+	10	_	24	210	260
+	3	_	90	860	NT
+	1	-	154	1290	NT
_	100	_	4	31	NT
+	_	3	6	40	80
+	_	1	6	40	NT
+	_	0.3	NT	150	250
+	_	0.1	NT	500	NT
_	_	3	NT	30	NT

<sup>&</sup>lt;sup>a</sup> C-26 IVX (2 x 10<sup>5</sup>/well) cells were cultured with IL-1 (10 pg/ml) and with increasing amounts of IL-1 RA or anti IL-1 R type I mAb (indicated in the table as nanograms per milliliter and micrograms per milliliter, respectively). At 24 and 48 h, culture supernatants were collected and subjected to IL-6 bioassay (in units per milliliter) or ELISA (in picograms per milliliter). Variation of triplicate determinations did not exceed 10%.
<sup>b</sup> Not tested.

Table III
Systemic administration of IL-1 RA and anti IL-1 R type I mAb does not improve weight loss<sup>a</sup>

Tumor Bearing	Treatment	No.	Host weight (g) on Day <sup>b</sup>			Tumor weight (g)b
			10	13	16	Day 16
+	PBS	6	25.0 ± 0.5	20.6 ± 1.0	19.0 ± 0.8	1.2 ± 0.2
+	IL-1RA	6	$25.5 \pm 0.7$	$23.0 \pm 0.9^{c}$	$21.0 \pm 1.0$	$1.1 \pm 0.3$
+	Anti IL-1R (type 1)	6	$25.9 \pm 0.6$	$20.1 \pm 0.8$	$19.1 \pm 0.9$	$1.4 \pm 0.5$
+	Rat IgG	5	$25.6 \pm 0.3$	$20.4 \pm 0.5$	$18.9 \pm 0.5$	$1.3 \pm 0.3$
_	PBS	5	$25.3 \pm 0.3$	$25.3 \pm 0.5$	$25.4 \pm 0.5$	

<sup>&</sup>lt;sup>a</sup> CD2F1 male mice were injected with C-26.IVX cells (day 0). On day 7 mice were randomized to receive PBS (0.25 ml, s.c. daily), IL-1RA (10 mg/kg, s.c. daily), anti-IL-1R type I mAb and polyclonal rat IgG (12.5 mg/kg, i.p. on days 7, 10, and 13).

Table IV Inhibition of Cachexia by Intratumoral Administration of IL-1RA is Dose Dependent<sup>a</sup>

Tumor Bearing	Treatment	Initial Weight (g)	Final Body Weight (g)	% Weight Loss	Tumor Weight (g)	Serum Glucose (mg/dl)
_	PBS	25.1 ± 0.9	25.8 ± 0.4		_	126 ± 3
+	PBS	$25.1 \pm 1.3$	$20.2 \pm 0.3$	19.5	$1.1 \pm 0.3$	$32 \pm 6$
+	IL-1RA (20)	$24.5 \pm 0.9$	$20.2 \pm 0.8$	17.5	$1.1 \pm 0.5$	NT <sup>b</sup>
+	IL-1RA (50)	$24.6 \pm 0.8$	$22.6 \pm 1.2^{c}$	8.2	$1.0 \pm 0.4$	$64 \pm 17^{d}$
+	IL-1RA (100)	$25.2 \pm 1.5$	$23.7 \pm 1.7^d$	5.9	$1.1 \pm 0.4$	$72 \pm 14^{d}$

<sup>&</sup>lt;sup>a</sup> CD2F<sub>1</sub> male mice were injected with C26.IVX cells (day 0). On day 7 mice were randomized. On day 9, 11, 13, and 14 tumors were injected with 0.1 ml PBS or with increasing amounts (shown in parentheses in micrograms) of IL-1RA. Mice were killed on day 15 and final body weight was determined after resection of tumor. Results are expressed as mean ± SD of five mice per group.

weight loss (both fat and muscle tissue), hypoglycemia, and serum IL-6 levels, were significantly improved by the treatment. Of note, the intratumoral administration of the 35F5 mAb or rat IgG at 100  $\mu$ g/tumor/injection, under the identical conditions described in Table IV, failed to improve cachexia in C-26 bearing mice (not shown). This observation can be explained by the fact that the IL-1RA is at

least two orders of magnitude more potent than the mAb in blocking IL-1 activities on the C-26 line in culture (Table II). It is also conceivable that the significantly lower m.w. of the IL-1RA—as compared with the mAb—allowed it to diffuse more easily and thus affect more cells when injected into the tumor.

The intratumoral administration of IL-1RA resulted in a

<sup>&</sup>lt;sup>b</sup> Results are expressed as weight of the host (total weight – estimated tumor weight) for days 10 and 13 and the weight of the host (following resection of the tumor) on day 16 ± SD.

c p < 0.025 from PBS-injected tumor-bearing mice.

<sup>&</sup>lt;sup>b</sup> Not tested.

<sup>&</sup>lt;sup>c</sup> Probability value of at least 0.03 from the group which received intratumoral injection of PBS.

<sup>&</sup>lt;sup>d</sup> Probability value of at least 0.01 from the group which received intratumoral injection of PBS.

Table V		
Reduction in cachetic parameter	rs by intratumoral	administration of IL-1RA <sup>a</sup>

Parameter	Group 1	Group 2	Probability Value	Group 3
Tumor Inoculation	+	+		_
Treatment	PBS	IL-1RA		PBS
Initial weight (g)	$25.7 \pm 1.0$	$26.0 \pm 1.5$	NS <sup>b</sup>	$26.6 \pm 0.8$
Final weight (g)	$21.5 \pm 1.4$	$24.6 \pm 1.2$	0.005	$27.7 \pm 0.8$
Host weight (g)	$20.3 \pm 1.4$	$23.4 \pm 1.2$	0.004	$27.7 \pm 0.8$
Tumor weight (g)	$1.19 \pm 0.05$	$1.13 \pm 0.12$	NS	
Epididymal fat (mg)	$102 \pm 57$	$204 \pm 74$	0.03	$310 \pm 102$
Dry weight (g)	$6.5 \pm 0.8$	$8.9 \pm 1.5$	0.03	10.2 ± 1.1
Serum glucose (mg/dl)	$57 \pm 12$	$105 \pm 12$	0.0004	$153 \pm 21$
Serum IL-6 (U/ml)	186 ± 24	$101 \pm 33$	0.03	$13 \pm 3$

<sup>&</sup>lt;sup>a</sup> CD2F<sub>1</sub> male mice were injected with C-26.IVX cells (day 0) and were randomized on day 7. On days 9, 11, 13, and 14 tumors were injected with PBS (0.2 ml, group 1) or IL-1RA (100 µg, group 2) and were killed on day 16. Mice in group 3 were non-tumor-bearing age-matched controls. Cachetic markers were quantified as described in *Materials and Methods*. Results are expressed as mean ± SD of six mice/group.

<sup>b</sup> Not significant.

relatively modest reduction (~40%) of serum IL-6, at a time when the protection of the host's weight was more pronounced (Table V). IL-1RA is a competitive antagonist to the action of IL-1 on the C-26 cell line (Table II). Its effect, therefore, is not permanent and may not be sufficient to inhibit IL-6 production by the additional tumor cells generated between the last administration of IL-1RA and the completion of the experiment. Alternatively, our still incomplete understanding of the regulation of IL-6R in the C-26 model, and the fact that serum IL-6 level may represent the unused portion of the cytokine in vivo, make it difficult to evaluate accurately the importance of serum IL-6 level. In addition, we cannot exclude the possibility that the level of IL-6 in the serum is of limited importance, as was recently demonstrated by May et al. (20)

To the best of our knowledge, the results presented here are the first to suggest that events occurring in the tumor mass contribute significantly to wasting, and that IL-1 plays an important role locally at the level of the tumor to initiate and/or exacerbate the wasting syndrome. IL-1 is a potent stimulator of IL-6 synthesis by the C-26 tumor line (10). The results shown here, although supporting the stimulator role of IL-1 in vivo, do not exclude the possibility that the cytokine may also amplify the production of a still undetermined factor/mediator that participates in C-26 cachexia, in addition to IL-6. These factors may also affect the physiology of tumor infiltrating macrophages present in C-26 tumors (10).

IL-6 is a multifunctional cytokine. In addition to its involvement in C-26-associated weight loss (9), it has been implicated in several unrelated chronic inflammatory conditions (21). Thus, an important question in the IL-6 field is how this pleiotropic cytokine contributes to such a wide variety of diseases and whether its involvement in vivo requires other factors that cooperate or synergize with its action.

The fact that intratumoral injection of IL-1RA did not influence tumor burden is important. A previous study at-

tempted to assess the effect of IL-1 on the development of cachexia in the MCG-101 tumor model (22). In that study, however, it was difficult to document a direct effect of IL-1 on wasting because the antibody used inhibited the tumor, thus implicating a role for the cytokine in promoting tumor growth.

The observation that systemically administered IL-1RA and 35F5 mAb could not affect cachexia is not surprising. Many conventional drugs and biologics, including cytokines and mAb, only minimally affect solid tumors, and several physiologic factors have been identified as responsible for the poor delivery of macromolecules to solid tumors (23). Identifying these factors in the C-26 model may further the design of therapeutics that inhibit cachexia.

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### References

- Langstein, H. N., and J. A. Norton. 1991. Mechanisms of cancer cachexia. Hematol. Oncol. Clin. North Am. 5:103.
- 2. Tisdale, M. J. 1991. Cancer cachexia. Br. J. Cancer 63:337.
- Warren, S. 1932. The immediate causes of death in cancer. Am. J. Med. Sci. 184:610.
- 4. van Eyes, J. 1982. Effect of nutritional status on response to therapy. *Cancer Res.* 42(Suppl.):747.
- Oliff, A., D. Defos-Jones, M. Boyer, D. Martinez, D. Kiefer, G. Vucolo, A. Wolfe, and S. Socher. 1987. Tumors secreting human TNF/cachectin induce cachexia in mice. Cell 50:555.
- Beutler, G., J. Mahoney, N. Letrang, P. Pekala, and A. Cerami. 1985. Purification of cachectin, a lipoprotein lipasesuppressing hormone from endotoxin induced RAW-2647 cells. J. Exp. Med. 161:984.
- Sherry, B. A., J. Gelin, Y. Fong, M. Marano, H. Wei, A. Cerami, S. F. Lowry, K. Lundholm, and L. L. Moldawer. 1989. Anticachectin/tumor necrosis factor antibodies attenuate development of cachexia in tumor models. FASEB J. 3:1956.
- 8. Tracey, K. J., H. Wei, K. R. Manogue, Y. Fong, D. G. Hesse, H. T. Nguyen, G. C. Kuo, B. Beutler, R. S. Cotran, A. Cerami,

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and S. F. Lowry. 1988. Cachectin/tumor necrosis factor induces cachexia anemia and inflammation. *J. Exp. Med. 167: 1211*.

- Strassmann, G., M. Fong, J. S. Kenney, and C. O. Jacob. 1992.
   Evidence for the involvement of IL-6 in experimental cancer cachexia. J. Clin. Invest. 89:1681.
- Strassmann, G., C. O. Jacob, R. Evans, D. Beall, and M. Fong. 1992. Mechanisms of experimental cancer cachexia. Interaction between mononuclear phagocytes and colon 26 carcinoma and its relevance to IL-6 mediated cancer cachexia. J. Immunol. 148:3674.
- Hannum, C. H., C. J. Wilcox, W. P. Arends, F. G. Joslin, D. J. Dripps, P. L. Heimdal, L. G. Armes, A. Sommer, S. P. Eisenberg, and R. C. Thompson. 1990. Interleukin 1 receptor antagonist activity of a human interleukin 1 inhibitor. *Nature* 343:336.
- Dinarello, C. A., and R. C. Thompson. 1991. Blocking IL-1: interleukin 1 receptor antagonist in vivo and in vitro. *Immunol. Today* 12:404.
- Eisenberg, S. P., R. J. Evans, W. P. Arend, E. Verderber, M. T. Brewer, C. H. Hannum, and R. C. Thompson. 1990. Primary structure and functional expression from complementary DNA of a human interleukin 1 receptor antagonist. *Nature* 343:341.
- 14. Chizzonite, R. A., T. Trivitt, P. L. Kilian, A. S. Stern, P. Numes, K. P. Parker, V. L. Kaffka, A. D. Chua, D. K. Lugg, and U. Gubbler. 1989. Two high affinity interleukin 1 receptors represents separate gene products. *Proc. Natl. Acad. Sci. USA* 86:8029.
- Tanaka, Y., H. Eda, T. Tanaka, T. Udagawa, T. Ishizaka, I. Horii, H. Ishitsuka, T. Kataoka, and T. Taguchi. 1990. Experimental cancer cachexia induced by transplantable colon 26 adenocarcinoma in mice. Cancer Res 50:2290.

- McCarthy, P. L., S. Abhayankar, S. Neben, G. Newman, C. Sieff, R. C. Thompson, S. J. Burakoff, and J. L. Ferrara. 1991.
   Inhibition of interleukin 1 by an interleukin 1 receptor antagonist prevents graft versus host disease. *Blood* 78:1915.
- McIntyre, K. W., G. J. Stepan, K. D. Kolinsky, W. R. Benjamin, J. L. Plocinski, K. L. Kaffka, C. A. Campen, R. A. Chizzonite, and P. L. Kilian. 1991. Inhibition of interleukin 1 binding and bioactivity in vitro and modulation of acute inflammation in vivo by IL-1 receptor antagonist and anti IL-1 receptor monoclonal antibody. J. Exp. Med. 173:931.
- Neta, R., J. J. Openheim, R. D. Schreiber, R. Chizzonite, G. D. Ledney, and T. J. MacVittie. 1991. Role of cytokines (interleukin 1, tumor necrosis factor and transforming growth factor) in natural and lipopolysaccharide enhanced radioresistance. J. Exp. Med. 173:1177.
- Gershenwald, J. E., Y. Fong, T. J. Fahey, S. E. Calvano, R. Chizzonite, P. L. Kilian, S. L. Lowry, and L. L. Moldawer. 1990. Interleukin 1 receptor blockade attenuates the host inflammatory response. *Proc. Natl. Acad. Sci. USA* 87:4966.
- May, L. T., H. Viguet, J. S. Kenney, N. Ida, A. C. Allison, and P. B. Shegal. 1992. High level of "complexed" interleukin 6 in human blood. J. Biol. Chem. 267:19698.
- 21. Akira, S., and T. Kishimoto. 1992. IL-6 and NF-IL6 in acute phase response and viral infection. *Immunol. Rev. 127:23*.
- Gelin, J., L. L. Moldawer, C. Lonnroth, B. Sherry, R. Chizzonite, and K. Lundholm. 1991. Role of endogenous tumor necrosis factor and interleukin 1 for experimental tumor growth and development of cancer cachexia. Cancer Res. 51:415.
- Jain, R. K. 1990. Vascular and interstitial barriers to the delivery of therapeutic agents in tumors. Cancer Metastasis Rev. 9:253.