

# Investigators' Brochure for the Clinical Trial of Photodynamic Therapy of Advanced Cancer

Cytoluminator Research Pty. Ltd.

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## List of Substances and Devices to be Trialled

**Photosensitising agent** aqueous solution of hydroxy-aluminium phthalocyanine disulphonate (abbreviated ALPcS<sub>2</sub>) for intravenous administration.

**Illuminator** a solid-state laser producing diffuse illumination from a hand-held light guide, at wavelengths predominantly between 684 – 686 nm, with variable power levels not exceeding 5 W, and with average intensities below the skin pain threshold (around 1 W/cm<sup>2</sup>).

**Fluorescence imager** a video camera with sensitivity to the infrared fluorescence produced by illuminated photosensitising agent (690 – 780 nm), but insensitive to the nonfluorescent and ambient light. An illumination accessory is incorporated for projecting light from the illuminator onto the subject with a uniform intensity distribution centred on the imager field of view.

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# Contents

|          |  |          |
|----------|--|----------|
| <b>1</b> | <b>Summary</b>   | <b>3</b> |
| <b>2</b> | <b>Introduction</b>  | <b>3</b> |
| <b>3</b> | <b>Physical, Chemical, Pharmaceutical Properties and Formulation</b> | <b>5</b> |
| <b>4</b> | <b>Nonclinical Studies</b>   | <b>6</b> |
| 4.1      | Nonclinical pharmacology . . . . .                                   | 6        |
| 4.2      | Pharmacokinetics and product metabolism in animals . . . . .         | 6        |
| 4.3      | Toxicology . . . . .   | 6        |
| 4.3.1    | Single dose . . . . .  | 6        |
| 4.3.2    | Repeated dose . . . . .  | 6        |
| 4.3.3    | Carcinogenicity . . . . .  | 6        |
| 4.3.4    | Special studies . . . . .  | 6        |
| 4.3.5    | Reproductive toxicity . . . . .                                      | 7        |
| 4.3.6    | Genotoxicity . . . . .   | 7        |
| <b>5</b> | <b>Effects in humans</b>   | <b>7</b> |
| 5.1      | Pharmacokinetics and product metabolism in humans . . . . .          | 7        |
| 5.2      | Safety and efficacy . . . . .  | 7        |
| 5.3      | Marketing experience . . . . .                                       | 7        |
| <b>6</b> | <b>Summary of data and guidance for the investigator</b>             | <b>7</b> |
|          | <b>References</b>  | <b>9</b> |

# 1 Summary

To be done.

## 2 Introduction

Photodynamic therapy (PDT), also known as photoradiation therapy, photochemotherapy, and by various other names, is the administration of a photosensitising agent followed by irradiation with light having the appropriate physical properties to activate the photosensitiser without heating the tissue [1, 2]. The photosensitiser is chosen to have minimal cytotoxicity when the body is kept in the dark, to be selectively taken up by the cancer cells and cleared from healthy tissue, and then to exhibit a potent cytotoxic effect under appropriate illumination. Due to the bimodal nature of the treatment, the practitioner is given increased flexibility and control as compared with traditional chemotherapy. For example, the concentration and localisation of sensitiser may be evaluated based on its fluorescence in real-time, and only once it has been adequately cleared from healthy tissue will the sensitiser be activated. There is an obvious diagnostic capability built into the treatment, for any region of increased fluorescence is very likely to contain cancer.

PDT has been in use worldwide for more than a decade, using a number of approved photosensitisers and illuminators. [3, 2]. Also see Refs. [4, 5].

Unlike chemotherapy or radiotherapy, PDT may be used repeatedly without long-term or cumulative effects [2], and unlike surgery or thermal ablation, there is little damage to underlying structures and the treated site heals normally with an “excellent cosmetic outcome” [1]. Although PDT should be performed early to achieve the best results, it is also ideally suited to palliation [2]. A number of cases have been reported in which PDT achieved complete control of post-salvage disease [1].

Existing procedures within the field of PDT mostly suffer from drawbacks that limit their usefulness to surface tumours and bodily fluids treated *ex vivo*, unless specialist equipment and/or surgical implants are used for internal light delivery. There are usually photosensitisation side-effects that discourage the widespread adoption of PDT. It is the object of this clinical trial to demonstrate the safety and efficacy of the photosensitisers and illuminators under development by Cytoluminator Research Pty. Ltd., an Australian company. Specifically, the trial is using a photosensitiser chosen from the class of *phthalocyanine* dyes, hydroxyaluminium phthalocyanine disulphonate, to be activated by light having most of its energy within the therapeutic wavelength interval of 684-686 nm (deep red). Unlike the most widely used porphyrin sensitisers, the phthalocyanines are relatively insensitive to ultraviolet light compared to the therapeutic wavelengths, greatly reducing the risks of inadvertent skin damage and enabling the participant to return to normal activity much sooner after systemic administration. This will facilitate the adoption of PDT into Australia, where high ultraviolet light levels have previously limited the use of systemic PDT to the most desperate cases, such as the work of researchers at the Royal Melbourne Hospital who achieved >25% long-term survival of patients with glioblastoma multiforme (a high grade brain cancer), using surgical resection followed by porphyrin PDT [6, 7, 8]. (It is well known that surgery followed by radio- and chemotherapy only achieves a two-year survival of around 5%.)

The choice of illumination wavelength is critical to the success of PDT. Whilst green, blue and ultraviolet light have negligible penetration beyond the skin and into the body, deep red and near infrared light can penetrate a significant distance (see Fig. 1). It is widely recognised that the heme molecule, found in hemoglobin and myoglobin, is responsible for most of the optical absorption of tissue, and that the optimal penetration depth is achieved for light having wavelengths around 700 – 800 nm. Unfortunately, none of the potential photosensitising agents that might be activated at these wavelengths have yet been found to possess the other necessary properties of a clinically relevant photosensitiser. It is less well known that deep red light around 685 nm can achieve close to the optimal penetration depth, and a potent photosensitiser at this wavelength is the object of the present study. The clinically relevant penetration depth is determined not only by the attenuation of light through tissue, but also by the minimum effective dose at the cancer and the maximum tolerated dose (MTD) at the skin. Through the use of a novel formulation (to be disclosed elsewhere), the sponsor has improved the tissue localisation of the phthalocyanine sensitiser to greatly reduce the photosensitivity of healthy tissue and thus extend the MTD and permit greater optical power densities without damaging the skin. Tumours may now be treated at a depth of several cm of tissue without significant

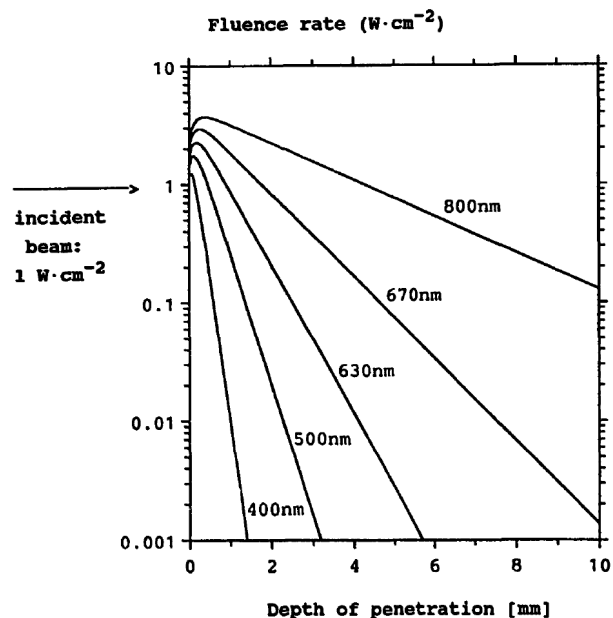


Figure 1: Light penetration into skin versus wavelength. Numerical modelling using the optical parameters of human skin, reproduced from [9].

pain, heating or damage to healthy structures. Due to excellent penetration of deep red light through bone, the sponsor has also achieved remissions of bony metastases and expects that it may eventually be possible to treat brain tumours using external illumination.

A widely accepted consequence of PDT is an immune response to cancer following treatment [10]. In animal models the immunity has been shown to last long after the original tumour was eliminated, since further cancer cell inoculations did not result in fresh tumours. Thus, PDT can be used to produce an in-situ vaccination against the cancer cell lines actually present within the participant. PDT has also been exploited *in vitro* to create inactivated cell vaccines against cancer [11], demonstrating that PDT-induced apoptosis followed by x-ray sterilisation is sufficient to produce the vaccination effect. Preliminary investigation by the sponsor has shown a strong immunological response, which may have contributed to the remission of remote metastases that were deeper than the presumed depth of light penetration. It is hoped that once the body is cleared of detectable tumours, the immune response will completely eradicate microscopic remnants and provide a true “cure”, meaning future relapse by the same cancer is very unlikely.

The scope of this therapy is very wide; any cancer is potentially treatable using the proposed sensitiser and sufficient illumination power and duration. It has been found that the present laser is sufficient to eradicate most breast tumours, as well as their metastases in the spine and other bones. Remission of multiple breast cancer metastases deep within the lung has also been demonstrated. Partial remission of prostate cancer has been demonstrated (after a single treatment only), and a partial remission has been achieved in a deep brain tumour (again after a single treatment). An area of present concern is tissue that is rich in blood, such as the liver and bone marrow, and it is unclear how far the light will penetrate. However, if a tumour is at least partially accessible to the illuminator, then the immune response may control the disease progression in those inaccessible regions of the body. (It is also possible to combine PDT with surgery to illuminate deeply buried tumours, with surgical debulking used where necessary.) In the case of leukemias, it may be possible to reduce the disease burden and stimulate an immune response, however no human studies have been performed by the sponsors at the time of writing. In a  $\sim 50$  kg dog with an unknown lymphoma occupying  $> 100$  cc of total lymphatic tumour volume,  $< 50\%$  of the tumour volume remained after three  $< 10$  min treatments on three consecutive days, suggesting that human lymphoma tumours will be susceptible to the present treatment.

Under the terms of the trial, the treatment protocol will be in accordance with the following summary. In a darkened room, a systemic administration of photosensitising agent shall be performed using either an intravenous bolus, or an intravenous infusion, with the measured dose of sensitiser filtered through a  $0.2 \mu\text{m}$  filter (or equivalent) prior to use. For an infusion, the sensitiser shall be injected into a bag of saline solution

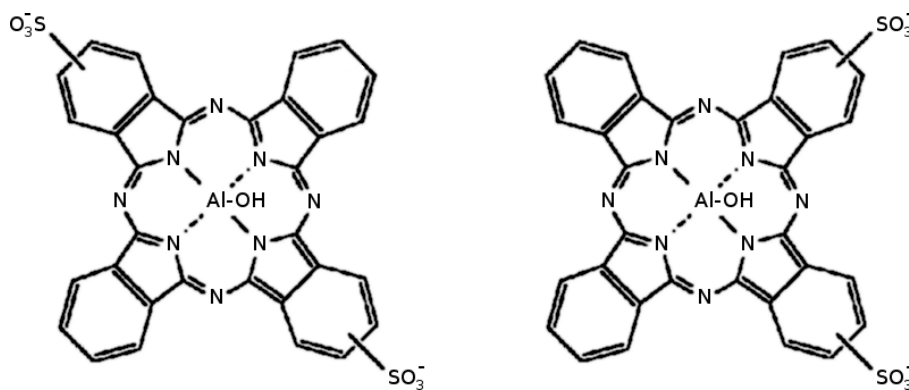


Figure 2: Chemical structures of hydroxy-aluminium phthalocyanine disulphonate. The structural isomers, differing only in the location of the sulphonyl functional groups on the phthalocyanine macrocycle, are prepared in a mixture to reduce intracellular dimerisation.

only. The rate at which the dilute sensitiser is infused will be limited by the practitioner so as to manage any discomfort and to allow for the fluorescence imager to highlight the localisation of the sensitiser into any tumour masses within its depth of observation. The amount of sensitiser present within the healthy tissue (the “background”) shall be measured from the intensity of the soft palate or the inner ear, at a known laser power output and distance between the imager and the patient. This shall be used as a reference for comparison with the same measurement taken over the following days, to a) verify the correct clearance from the healthy tissue before commencement of tumour irradiation, and b) determine when it is safe for the patient to begin an escalation of sun exposure to normal levels. The patient shall be made aware of the requirement that they avoid sun exposure until the practitioner has given them approval. This is expected to be around one week from treatment.

### 3 Physical, Chemical, Pharmaceutical Properties and Formulation

The photosensitising agent to be used, as prepared by the sponsor, is an aqueous solution of the sodium salt of hydroxy-aluminium phthalocyanine disulphonate,  $\text{Al}(\text{OH})\text{Pc}(\text{SO}_3^- \text{Na}^+)_2$  (abbreviated  $\text{AlPcS}_2$ ). The pH is raised by  $\text{NaOH}$  to minimise aggregation of the molecules, as under acidic or neutral conditions the  $\text{AlPcS}_2$  molecules will tend to dimerise and lose their photodynamic activity [12, 13]. The chemical structures of the two isomers of  $\text{AlPcS}_2$  are depicted in Fig. 2. The preparation to be trialled is a mixture of the two, which has been shown to exhibit greatly reduced dimerisation over the use of one isomer or another [12], both in the as-supplied concentrated solution and in the intracellular compartment (where concentrations are much higher than in the serum).

$\text{AlPcS}_2$  is structurally similar to the other phthalocyanines, and exhibits similar physical and chemical properties. The solution is translucent and cyan (blue-green) in colour, due to the strong absorption of red light. It is relatively photostable. When illuminated with UV-A or red light, the sensitiser absorbs light energy and fluoresces in the deep red part of the spectrum, with a peak emission wavelength around 670 – 690 nm [13] as well as a significant amount of near-IR emission extending to 780 nm. It also efficiently promotes  $\text{O}_2$  molecules to a singlet excited state [13], which contains excess energy and is highly reactive. Singlet oxygen, and the reactive oxygen species (ROS) it produces, are highly cytotoxic but have a very short lifetime and a correspondingly short range of diffusion ( $< 0.02 \mu\text{m}$ ). For this reason, the photodynamic cytotoxicity is entirely confined to the cells in which the sensitiser is localised at the time of illumination. The sensitiser is not consumed in the process.

Pure  $\text{AlPc}$  is known to be thermally stable up to its boiling point *in vacuo* [14], and the sponsor has found its sulphonated derivatives to be similarly stable when dried at 250 °C. The addition of sulphonyl functional groups results in water soluble forms, with pharmacokinetic properties dependent on the degree of sulphonation and thus the net ionic charge and hydrophilicity or lipophilicity of the molecule. It is known that  $\text{AlPc}$  and its monosulphonate derivative ( $\text{AlPcS}_1$ ) will accumulate in the skin and eyes for extended periods, due to lipophilicity (and poor water solubility). The tetrasulphonate ( $\text{AlPcS}_4$ ) largely remains

in the interstitial compartment, due to its hydrophilicity. These substances are detrimental to the light penetration and increase the phototoxicity to healthy tissue. The disulphonate (AlPcS<sub>2</sub>) and trisulphonate (AlPcS<sub>3</sub>) are taken up specifically by cancer cells and immune cells, facilitated by endocytosis of low-density lipoproteins which contain bound sensitiser. AlPcS<sub>2</sub> exhibits the greatest therapeutic effect by localising to the mitochondrial membrane upon illumination, inactivating many proteins anchored there. The main therapeutic target has been identified as the anti-apoptotic proteins Bcl-2 and its relatives, depletion of which triggers apoptosis through loss of mitochondrial membrane potential and release of cytochrome-c from the mitochondria. (Bcl-2 is part of the signalling pathway used by cytotoxic T cells and natural killer cells to induce apoptosis in cancer and virally infected cells.) Even cells that are unable to complete the apoptotic process will die due to loss of mitochondrial function. AlPcS<sub>3</sub> may prove beneficial in subsequent sensitiser mixtures, possibly due to reduction of intracellular aggregation, which will be investigated further.

It is anticipated that the highest administered dose will be 1.0 mg/kg (of body mass). The sensitiser will be supplied in a sterile, pre-filtered solution at a concentration of 1.0 mg/ml, packaged in a glass bottle with a rubber septum for withdrawal into a syringe. The solution should be stored in a cool, dark place (preferably in a refrigerator), although it is not known whether this is necessary. Due to the possibility of aggregation and settling (which has not been ruled out), the bottle shall be shaken vigorously prior to use, to ensure the correct dose is obtained. It shall be injected into isotonic saline via an in-line 0.2  $\mu$ m filter. As the concentrated solution is not isotonic, is not suitable for direct injection and must be diluted into saline before administration. Only saline shall be used, as other solutions may promote dimerisation or aggregation after filtration. For intratumoural injection or intravenous bolus, the sponsor is investigating the stability of an as-supplied isotonic sensitiser solution.

## 4 Nonclinical Studies

### 4.1 Nonclinical pharmacology

Ref. [15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 5].

### 4.2 Pharmacokinetics and product metabolism in animals

Ref. [25, 26, 27, 28, 15, 29, 30, 31, 32, 33, 34, 35, 36, 37, 11, 38]. (For animal models of bone metastases treated using PDT, see Ref. [39].)

For the correct wavelength see Ref. [40, 41].

### 4.3 Toxicology

#### 4.3.1 Single dose

#### 4.3.2 Repeated dose

Ref. [42, 43].

#### 4.3.3 Carcinogenicity

None reported.

#### 4.3.4 Special studies

N/A

#### **4.3.5 Reproductive toxicity**

No studies have been performed.

#### **4.3.6 Genotoxicity**

No studies have been performed.

## **5 Effects in humans**

### **5.1 Pharmacokinetics and product metabolism in humans**

A functionalised silicon phthalocyanine, referred to as Pc4, has been investigated by researchers from Case Western Reserve University for more than a decade, see Ref. [5]. It has entered early clinical trials.

### **5.2 Safety and efficacy**

Mixtures of sulphonated aluminium phthalocyanines have been used to treat more than 100 cases of metastatic breast cancer in a Phase 3 clinical trial in Russia, see Refs. [44, 45]. This provides strong support for the safety of the present study, and the sponsor believes the Australian-made sensitiser to be of higher purity than the Russian version, which is poorly defined. The cited trial made use of the clinically ineffective wavelength of 673 nm at a much higher fluence for skin metastases, while resorting to implantation of interstitial fibre-optics to illuminate deep tumours.

### **5.3 Marketing experience**

The sponsor has no marketing experience with this formulation.

## **6 Summary of data and guidance for the investigator**

The photosensitiser is expected to remain within tumours for extended periods (days or weeks) until they have been successfully killed by the illuminator. This allows the course of irradiation to be extended over several days without further administration of sensitiser. In fact, this may be the key to achieving complete remission even when the tumour appears to be completely eradicated. Viable cancer cells could remain, due to insufficient light penetration through the tumour mass, inadequate oxygen perfusion during the treatment, or inhomogeneous sensitiser uptake. In addition, a more powerful immune response could result from ongoing exposure of dying cancer cells to the immune system, when compared to simultaneous destruction of the entire tumour. This will be a topic of investigation for the trial, along with the use of immune modulating adjuvants such as: granulocyte colony stimulating factor (G-CSF, marketed as Filgrastim), a cytokine which boosts the blood neutrophil count and level of activity; or granulocyte-monocyte colony stimulating factor (GM-CSF), a cytokine which boosts the neutrophil and macrophage counts and level of activity.

It has been found by the sponsor, that irradiation is painless and only a mild warmth is felt, however a sharp pain response is usually felt after an excessive duration of tumour irradiation, probably due to release of sensitiser into nearby tissue by necrotic cells. This occurs after around 10 – 15 min of illumination at 3 W laser power onto a tumour within intact breast tissue or axillary lymph nodes. After this time, the region of the tumour is very sensitive to light from the illuminator, and it is obvious that a change has occurred. Further irradiation would not be possible without anaesthesia, nor does it appear beneficial since apoptotic cell death occurs at a small fraction of this dose. When the same region is observed on the following day, the tumours that have been irradiated no longer fluoresce above the background level. It is therefore straightforward to detect any tumours that have been missed and illuminate them again. Once there are no discernable bright regions on the fluorescence image, the practitioner must be guided by other sources of

information to illuminate the deeper tumours, such as MRI, CT or PET scans, in the hopes that these may be treated with the present level of illumination (and a longer duration of exposure). It is expected that a surface tumour, such as just below the skin, will require at most  $2 \text{ s/cm}^2$  (of skin surface area), to achieve apoptotic cell death with the present 5 W illuminator. It is unknown how the dose must be scaled for very deep tumours as this is a topic of further research – if the practitioner is willing to perform a few hours of illumination, it is possible to greatly reduce the tumour burden of even widespread metastatic cancer for which patient death is imminent. For that reason, use in palliative care of post-salvage patients is strongly encouraged. In one case of widely metastatic breast cancer in which multiple treatments were performed along with immune modulating adjuvants, near complete remission was achieved over a period of less than one year of continuous improvement.

## Revsion History

**DRAFT 28 Dec 2008** Initial version by Darren Freeman



## References

- [1] Colin Hopper. Photodynamic therapy: a clinical reality in the treatment of cancer. *Lancet Oncol.*, 1(4):212–219, Dec 2000.
- [2] Stanley B Brown, Elizabeth A Brown, Ian Walker. The present and future role of photodynamic therapy in cancer treatment. *Lancet Oncol.*, 5(8):497–508, Aug 2004.
- [3] Massimo Conio, Alan J Cameron, Amitabh Chak, Sabrina Bianchi, Rosangela Filiberti. Endoscopic treatment of high-grade dysplasia and early cancer in Barrett’s oesophagus. *Lancet Oncol.*, 6:311–321, May 2005.
- [4] Thomas J Dougherty, Charles J Gomer, Barbara W Henderson, Giulio Jori, David Kessel, Mladen Korbelik, Johan Moan, Qian Peng. Photodynamic therapy. *J. Natl. Cancer Inst.*, 90(12):889–905, Jun 1998.
- [5] Janine D Miller, Elma D Baron, Heather Scull, Andrew Hsia, Jeffrey C Berlin, Thomas McCormick, Valdir Colussi, Malcolm E Kenney, Kevin D Cooper, Nancy L Oleinick. Phtodynamic therapy with the phthalocyanine photosensitizer Pc 4: The case experience with preclinical mechanistic and early clinical-translational studies. *Toxicol. and Appl. Pharmacol.*, 224:290–299, Feb 2007.
- [6] Stanley S Stylli, Andrew H Kaye, Lachlan MacGregor, Megan Howes, Priya Rajendra. Photodynamic therapy of high grade glioma - long term survival. *J. Clin. Neurosci.*, 12(4):389–398, May 2005.
- [7] Stanley S Stylli, Andrew H Kaye. Photodynamic therapy of cerebral glioma - a review: Part I - a biological basis. *J. Clin. Neurosci.*, 13(6):615–625, Jul 2006.
- [8] Stanley S Stylli, Andrew H Kaye. Photodynamic therapy of cerebral glioma - a review: Part II - clinical studies. *J. Clin. Neurosci.*, 13(7):709–717, Aug 2006.
- [9] Martin Ochsner. Light scattering of human skin: a comparison between zinc(II)-phthalocyanine and Photofrin II(R). *J. Photochem. Photobiol. B: Biol.*, 32(1-2):3–9, Jan 1996.
- [10] Frederieke H van Duijnhoven, Remco I J M Aalbers, Jeroen P Rovers, Onno T Terpstra, Peter J K Kuppen. The immunological consequences of photodynamic treatment of cancer, a literature review. *Immunobiol.*, 207:105–113, 2003.
- [11] M Korbelik, B Stott, J Sun. Photodynamic therapy-generated vaccines: relevance of tumor cell death expression. *Brit. J. Cancer*, 97:1381–1387, 2007.
- [12] R Edrei, V Gottfried, J E Van Lier, S Kimel. Sulfonated phthalocyanines: photophysical properties, in vitro cell uptake and structure-activity relationships. *J. Porphyrins Phthalocyanines*, 2:191–199, 1998.
- [13] Krystyna Palewska, Marta Sujka, Barbara Urasińska-Wójcik, Juliusz Sworakowski, Józef Lipiński, Stanislav Nešpurek, Jan Rakušan, Marie Karásková. Light-induced effects in sulfonated aluminum phthalocyanines - potential photosensitizers in the photodynamic therapy: Spectroscopic and kinetic study. *J. Photochem. Photobiol. A: Chem.*, 197(1):1–12, Jun 2008.
- [14] M E Azim-Araghi, A Krier. The influence of ammonia, chlorine and nitrogen dioxide on chloro-aluminium phthalocyanine thin films. *Appl. Surf. Sci.*, 119(3-4):260–266, Oct 1997.
- [15] S S Stylli, J S Hill, W H Sawyer, A H Kaye. Phthalocyanine photosensitizers for the treatment of brain tumours. *J. Clin. Neurosci.*, 2(1):64–72, 1995.
- [16] G M Malham, R J Thomsen, G J Finlay, B C Baguley. Subcellular distribution and photocytotoxicity of aluminium phthalocyanines and haematoporphyrin derivative in cultured human meningioma cells. *Brit. J. Neurosurgery*, 10(1):51–57, Feb 1996.
- [17] Yu Luo, David Kessel. Initiation of apoptosis versus necrosis by photodynamic therapy with chloroaluminium phthalocyanine. *Photochem. Photobiol.*, 66(4):479–483, Oct 1997.
- [18] Hyeong-Reh Choi Kim, Yu Luo, Gangyong Li, David Kessel. Enhanced apoptotic response to photodynamic therapy after bcl-2 transfection. *Cancer Res.*, 59(14):3429–3432, Jul 1999.

- [19] J Usuda, Song-mao Chiu, Erin S Murphy, Minh Lam, Anna-Liisa Nieminen. Domain-dependent photo-damage to Bcl-2: a membrane-anchorage region is needed to form the target of phthalocyanine photosensitization. *J. Bio. Chem.*, 278(3):2021–2029, Jan 2003.
- [20] Liang-yan Xue, Song-mao Chiu, Aline Fiebig, David W Andrews, Nancy L Oleinick. Photodamage to multiple Bcl-xL isoforms by photodynamic therapy with the phthalocyanine photosensitizer Pc 4. *Oncogene*, 22(58):9197–9204, Dec 2003.
- [21] S-M Chiu, L-Y Xue, K Azizuddin, N L Oleinick. Photodynamic therapy-induced death of HCT 116 cells: apoptosis with or without Bax expression. *Apoptosis*, 10(6):1357–1368, Dec 2005.
- [22] Hui-Fang Huang, Yuan-Zhong Chen, Yong Wu. ZnPcS<sub>2</sub>P<sub>2</sub>-based photodynamic therapy induces mitochondria-dependent apoptosis in K562 cells. *Acta Biochimica et Biophysica Sinica*, 37(7):488–494, 2005.
- [23] Liang-yan Xue, Song-mao Chiu, Nancy L Oleinick. Differential responses of Mcl-1 in photosensitized epithelial vs lymphoid-derived human cancer cells. *Oncogene*, 24(46):6987–6992, Oct 2005.
- [24] H Kolarova, P Nevrelva, R Bajgar, D Jirova, K Kejlova, M Strnad. In vitro photodynamic therapy on melanoma cell lines with phthalocyanine. *Toxicol. in Vitro*, 21(2):249–253, Mar 2007.
- [25] R Cubeddu, R Ramponi, P Taroni G Canti. Time-gated fluorescence spectroscopy of porphyrin derivatives and aluminium phthalocyanine incorporated in vivo in a murine ascitic tumour model. *J. Photochem. Photobiol. B: Biol.*, 11(3-4):319–328, Dec 1991.
- [26] M Shopova, V Mantareva, K Krastev, D Hadjiolov, A Milev, K Spirov, G Jori, F Ricchelli. Comparative pharmacokinetic and photodynamic studies with zinc(II) phthalocyanine in hamsters bearing an induced or transplanted rhabdomyosarcoma. *J. Photochem. Photobiol. B: Biol.*, 16(1):83–89, Oct 1992.
- [27] Ji-Yao Chen, Wen Chen, Huai-Xin Cai, Rong-Chun Dong. Studies on pharmacokinetics of sulfonated aluminum phthalocyanine in a transplantable mouse tumor by *in vivo* fluorescence. *J. Photochem. Photobiol. B: Biol.*, 18(2-3):233–237, May 1993.
- [28] George M Peavy, Tatiana B Krasieva, Bruce J Tromberg, E Dave Eusantos, Michael W Berns. Variation in the distribution of a phthalocyanine photosensitizer in naturally occurring tumors of animals. *J. Porphyrins Phthalocyanines*, 27(3):271–277, Mar 1995.
- [29] S S Stylli, J S Hill, W H Sawyer, A H Kaye. Aluminium phthalocyanine mediated photodynamic therapy in experimental malignant glioma. *J. Clin. Neurosci.*, 2(2):146–151, April 1995.
- [30] Natalia Kazachkina, Natalia Zharkova, Galina Fomina, Raisa Yakubovskaya, Victor Sokolov, Eugeniji Luk’yanetz. Pharmacokinetical study of Al- and Zn-sulphonated phthalocyanines. *Proc. SPIE*, 2924:233–242, Sep 1996.
- [31] W-S Chan, N Bresseur, C La Madeleine, R Ouellet, J E van Lier. Efficacy and mechanism of aluminium phthalocyanine and its sulphonated derivatives mediated photodynamic therapy on murine tumours. *European J. Cancer*, 33(11):1855–1859, 1997.
- [32] Gianfranco Canti, Angelo Nicolin, Rinaldo Cubeddu, Paola Taroni, Gaetano Bandieramonte, Gianluca Valentini. Antitumor efficacy of the combination of photodynamic therapy and chemotherapy in murine tumors. *Cancer Lett.*, 125(1-2):39–44, 1998.
- [33] Merrill J Egorin, Eleanor G Zuhowski, Dorothy L Sentz, Jason M Dobson, Patrick S Callery, Julie L Eiseman. Plasma pharmacokinetics and tissue distribution in CD<sub>2</sub>F<sub>1</sub> mice of Pc4 (NSC 676418), a silicone phthalocyanine photodynamic sensitizing agent. *Cancer Chemother. Pharmacol.*, 44(4):283–294, Oct 1999.
- [34] Katrin Kalka, Nihal Ahmad, Tracy Criswell, David Boothman, Hasan Mukhtar. Up-regulation of clusterin during phthalocyanine 4 photodynamic therapy-mediated apoptosis of tumor cells and ablation of mouse skin tumors. *Cancer Res.*, 60:5984–5987, 2000.
- [35] Cecilia M Whitacre, Taroh H Satoh, Liang yan Xue, Nahida H Gordon, Nancy L Oleinick. Photodynamic therapy of human breast cancer xenografts lacking caspase-3. *Cancer Lett.*, 179(1):43–49, May 2002.

- [36] John E George, Yusra Ahmad, Davood Varghai, Xiaolin Li, Jeffrey Berlin, David Jackowe, Marc Jungermann, Michael S Wolfe, Lothar Lilge, Ali Totonchi, Rachel L Morris, Allyn Peterson, W David Lust, Malcolm E Kenney, Charles L Hoppel, Jiayang Sun, Nancy L Oleinick, David Dean. Pc 4 photodynamic therapy of U87-derived human glioma in the nude rat. *Lasers Surg. Med.*, 36(5):383–389, Jun 2005.
- [37] Antonella Borgatti-Jeffreys, Stephen B Hooser, Margaret A Miller, Michael D Lucroy. Phase I clinical trial of the use of zinc phthalocyanine tetrasulfonate as a photosensitizer for photodynamic therapy in dogs. *Am. J. Vet. Res.*, 68(4):399–404, Apr 2007.
- [38] Yan Huang, Guoxing Xu, Yiru Peng, Hong Lin, Xuedong Zheng, Maosong Xie. Zinc phthalocyanine tetrasulfonate (ZnPcS<sub>4</sub>): a new photosensitizer for photodynamic therapy in choroidal neovascularization. *J. Ocular Pharmacol. Therapeutics*, 23(4):377–386, 2007.
- [39] S Burch, S K Bisland, J Siewerdsen, A Bogaards, D Moseley, A Yee, J Finkelstein, B Wilson. Photodynamic therapy for the treatment of metastatic lesions in bone: Studies in rat and porcine models. *Proc. SPIE*, 5315:76–87, Jan 2004.
- [40] Claes af Klinteberg, Antonio Pifferi, Stefan Andersson-Engels, Rinaldo Cubeddu, Sune Svanberg. In vivo absorption spectroscopy of tumor sensitizers with femtosecond white light. *Appl. Opt.*, 44(11):2213–2220, Apr 2005.
- [41] John Griffiths, Janet Cruse-Sawyer, Simon R Wood, Jack Schofield, Stanley B Brown, Bryan Dixon. On the photodynamic therapy action spectrum of zinc phthalocyanine tetrasulphonic acid *in vivo*. *J. Photochem. Photobiol. B: Biol.*, 24(1):195–199, Apr 1994.
- [42] Wei Liu, Naisheng Chen, Hongtao Jin, Jinling Huang, Jinfeng Wei, Jie Bao, Chenghe Li, Yan Liu, Xueyong Li, Aiping Wang. Intravenous repeated-dose toxicity study of ZnPcS<sub>2</sub>P<sub>2</sub>-based-photodynamic therapy in beagle dogs. *Regul. Toxicol. Pharmacol.*, 47(3):221–231, Apr 2007.
- [43] Zibo Zhang, Hongtao Jin, Jie Bao, Fang Fang, Jinfeng Wei, Aiping Wang. Intravenous repeated-dose toxicity study of ZnPcS<sub>2</sub>P<sub>2</sub>-based-photodynamic therapy in Wistar rats. *Photochem. Photobiol. Sci.*, 5(11):1006–1017, Nov 2006.
- [44] Elena G Vakulovskaya, Victor V Shental, Yury V Buidenok, Georgui I Solomakho, Tatiana M Nadezhdina. Photodynamic therapy of breast cancer with photosense. *Proc. SPIE*, 4248:46–48, Jan 2001.
- [45] Elena G Vakulovskaya, Victor P Letyagin, Loubov V Umnova, Georgiu N Vorozhcsov, Victor Philinov. Photodynamic therapy and fluorescent diagnostics of breast cancer. *Proc. SPIE*, 5315:152–156, Jan 2004.